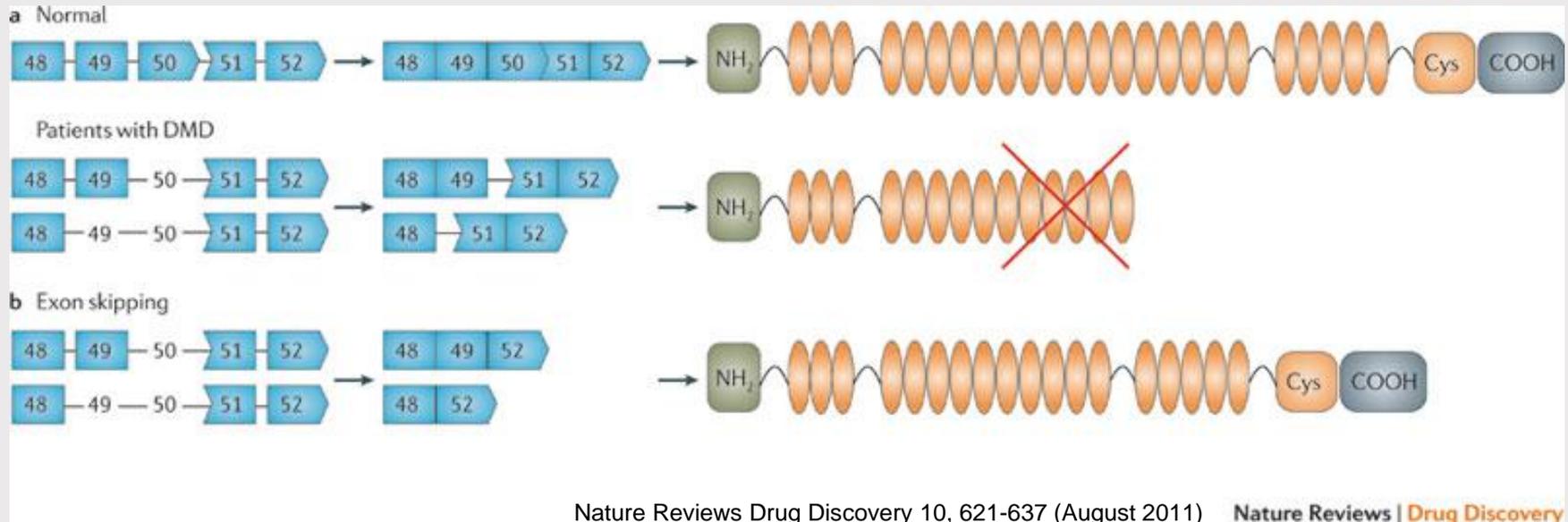


Duchenne Muscular Dystrophy

- Duchenne muscular dystrophy (DMD) is the most common form of muscular dystrophy, occurring in approximately 1 in 3,500.
- Promising new treatment to restore partially functional dystrophin protein by exon skipping is currently in clinical trials



Exon Skipping



Skip out of frame exons to put reading frame back in frame, resulting in a shorter Becker-like transcript

Monitoring response to exon skipping therapy

- 6 minute walk--most widely used clinical endpoint
 - Subject to high variability
 - May take up to a year to show improvement
 - More patients required to power statistics
 - Not applicable for wheelchair bound patients
 - Skipped mRNA product quantitation
 - Indirect measurement
 - mRNA level do not always correlate to protein levels
 - Dystrophin protein by immunohistochemistry or Western blot
 - Indirect measurement
 - Highly variable
 - Does not detect splice isoforms
- Longer trials and increased costs

Dystrophin Quantitation

Goal is to use Dystrophin protein as an 'efficacy surrogate endpoint'.

- Dystrophin is a clinically relevant endpoint measurement
- Dystrophin is a low abundant protein, accounting for only 0.002% of total striated muscle
- Rapid analysis will aid critical Phase 2/3 assessments
- Need for a rapid, sensitive and reliable assay to quantitate dystrophin

Dystrophin Quantitation

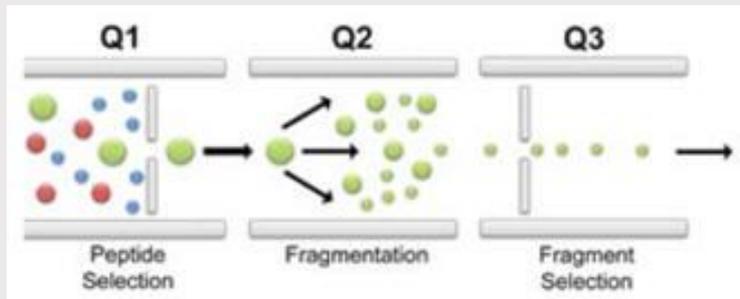
EMA suggestion:

Convene international consensus working group to establish standards in dystrophin protein as endpoint

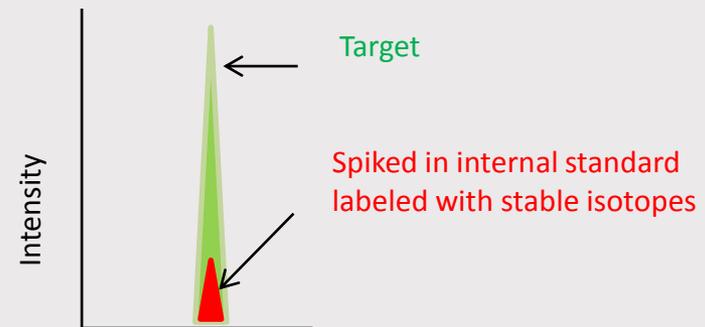
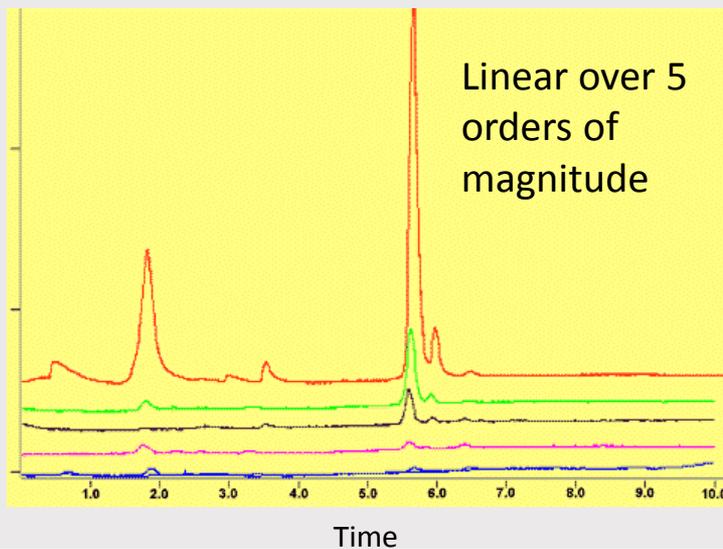
- Francesco Muntoni (London); Eric Hoffman (DC).
 - Call for participating testing labs
 - 7 labs volunteered
 - 1st Meeting to discuss approach
 - Distribution of biopsies, blinded
 - Test minimum of 3 replicates
 - Send data to central statistician
 - Intra-rater, inter-rater reliability defined
 - 2nd Meeting to discuss results

Immunofluorescence
Immunoblotting
- None reliable

Need new assay: MRM for targeted quantitative proteomics and validation

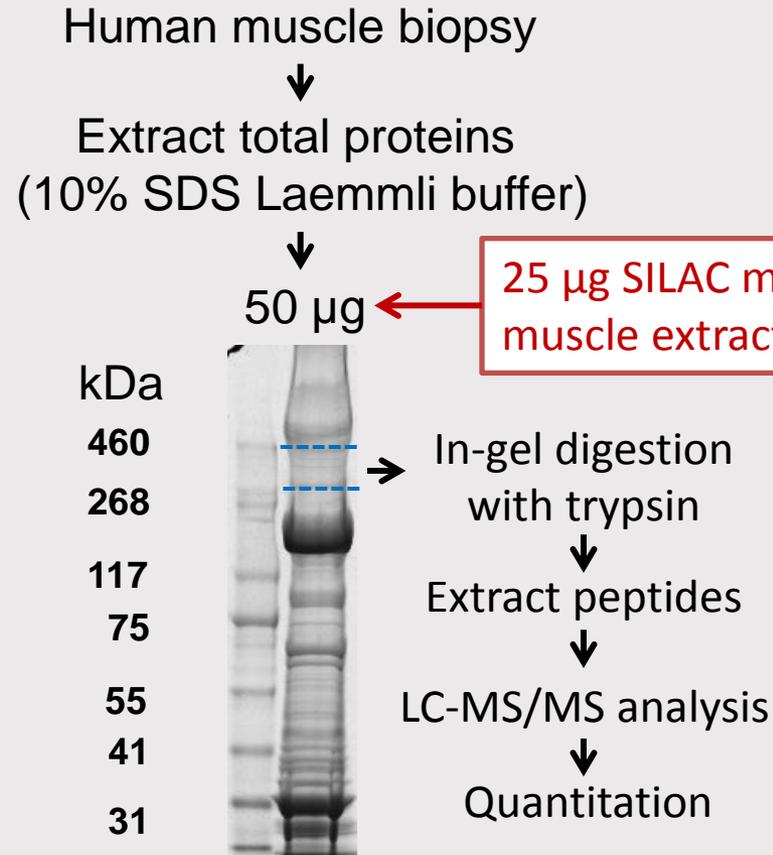


Multiple reaction monitoring is used to multiplex validation of selected targets in serial biological samples as an improved alternative to western-blot or ELISA. The FDA approve use as tool to measure surrogate biomarkers in a number of diseases.



Targeted MS Quantification of Dystrophin and other proteins in Human Muscle Biopsies

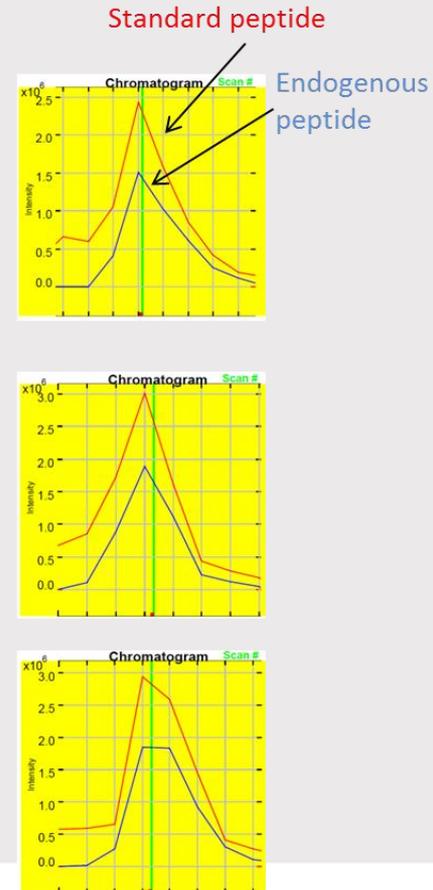
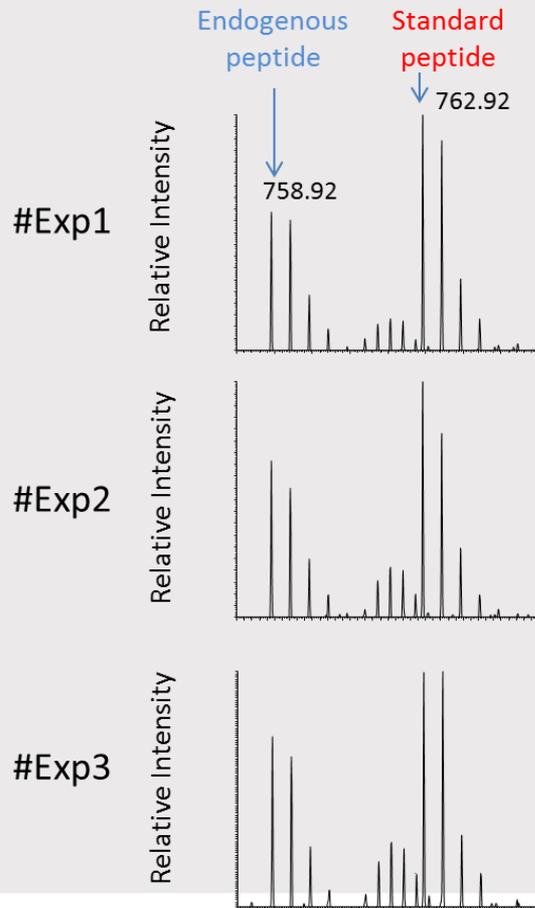
- Mass Spectrometry based quantitation requires a stable isotope standard.
- We use labeled mouse muscle to serve as the labeled dystrophin.
- This “heavy” SILAC mouse was generated in-house by maintaining a colony on $^{13}\text{C}_6$ -Lysine or ^{15}N feed.



Reproducibility of the method performed on muscle samples from a healthy donor (< 7% CV)

MS

Extracted ion chromatogram



Dystrophin Peptides Targeted by MS/MS

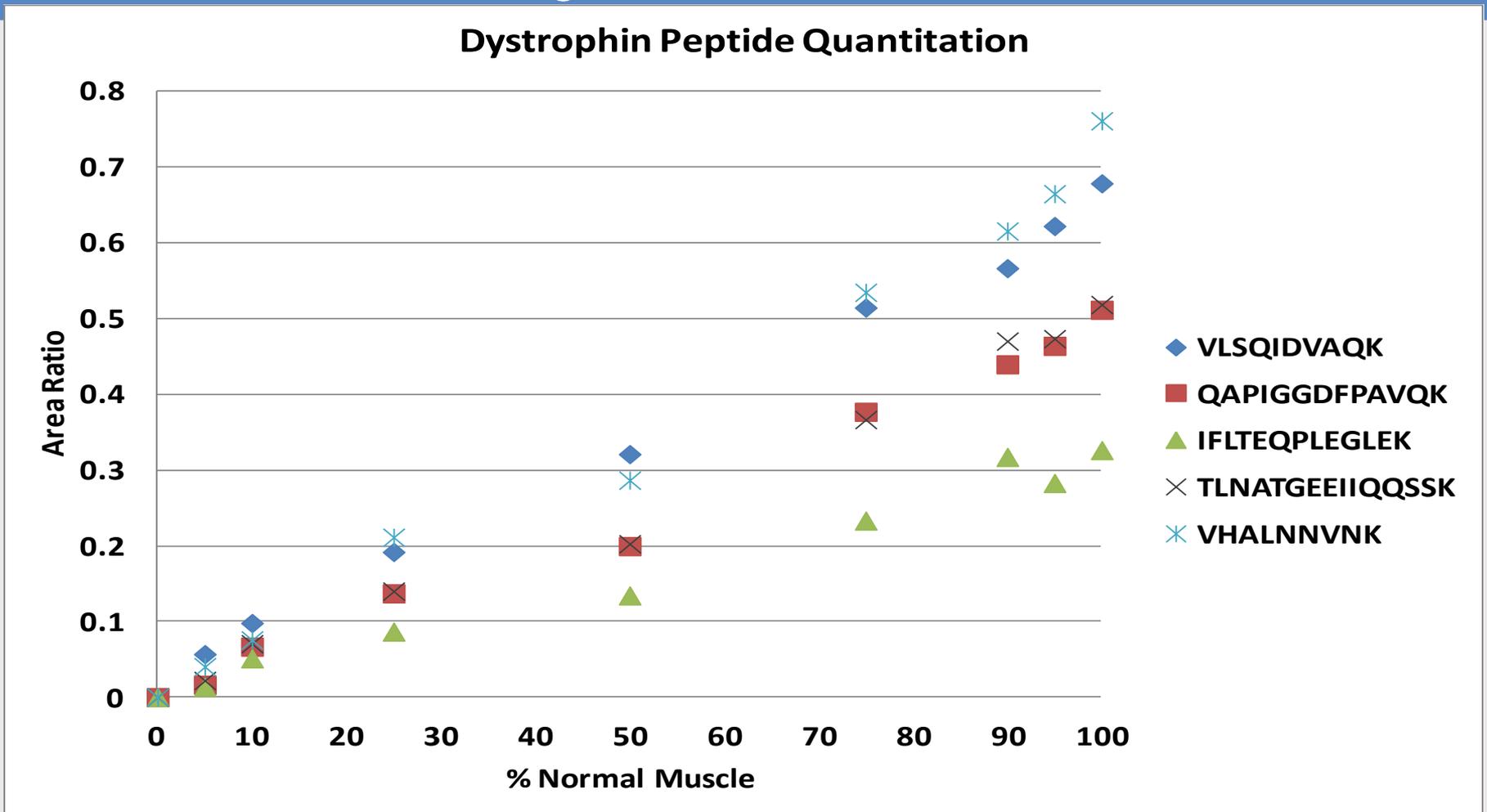
| >gil5032303ref NP_004009.1 dystrophin Dp71ab isoform [Homo sapiens] | |
|--|--|
| DAELIAEAK | |
| 480.2558->418.2291 | |
| 480.2558->531.3131 | |
| 480.2558->644.3972 | |
| 480.2558->773.4398 | |
| DAELIAEAK[13C6] | |
| 483.2659->424.2493 | |
| 483.2659->537.3333 | |
| 483.2659->650.4174 | |
| 483.2659->779.4600 | |
| LRQLLEQPQAEAK | |
| 508.6228->418.2291 | |
| 508.6228->546.2877 | |
| 508.6228->643.3404 | |
| 508.6228->771.3990 | |
| 508.6228->507.2664 | |
| LRQLLEQPQAEAK[13C6] | |
| 510.6295->424.2493 | |
| 510.6295->552.3079 | |
| 510.6295->649.3606 | |
| 510.6295->777.4192 | |
| 510.6295->510.2766 | |

| IFLTEQPLEGLEK | |
|-----------------------|--|
| 758.9165->446.2604 | |
| 758.9165->785.4398 | |
| 758.9165->1042.5410 | |
| 758.9165->1143.5886 | |
| 758.9165->1256.6727 | |
| IFLTEQPLEGLEK[13C6] | |
| 761.9266->452.2806 | |
| 761.9266->791.4600 | |
| 761.9266->1048.5612 | |
| 761.9266->1149.6089 | |
| 761.9266->1262.6929 | |
| QAPIGGDFPAVQK | |
| 664.3538->542.3291 | |
| 664.3538->689.3975 | |
| 664.3538->918.4674 | |
| 664.3538->1031.5515 | |
| 664.3538->1128.6042 | |
| QAPIGGDFPAVQK[13C6] | |
| 667.3639->548.3493 | |
| 667.3639->695.4177 | |
| 667.3639->924.4876 | |
| 667.3639->1037.5717 | |
| 667.3639->1134.6244 | |
| TLNATGEEIIQQSSK | |
| 809.9177->577.2935 | |
| 809.9177->690.3775 | |
| 809.9177->932.5042 | |
| 809.9177->1118.5682 | |
| 809.9177->1219.6160 | |
| TLNATGEEIIQQSSK[13C6] | |
| 812.9278->583.3137 | |
| 812.9278->696.3977 | |
| 812.9278->938.5244 | |
| 812.9278->1124.5885 | |
| 812.9278->1225.6361 | |

| VLSQIDVAQK | |
|-----------------------------|--|
| 550.8191->445.2764 | |
| 550.8191->560.3033 | |
| 550.8191->673.3874 | |
| 550.8191->801.4459 | |
| 550.8191->888.4780 | |
| VLSQIDVAQK[13C6] | |
| 553.8292->451.2966 | |
| 553.8292->566.3235 | |
| 553.8292->679.4076 | |
| 553.8292->807.4661 | |
| 553.8292->894.4982 | |
| LLDLLEGLTGQKLPK | |
| 546.6660->771.4717 | |
| 546.6660->941.5773 | |
| 546.6660->1070.6199 | |
| 546.6660->592.3556 | |
| 546.6660->706.4111 | |
| LLDLLEGLTGQK[13C6]LPK[13C6] | |
| 550.6795->783.5121 | |
| 550.6795->953.6177 | |
| 550.6795->1082.6603 | |
| 550.6795->598.3758 | |
| 550.6795->712.4313 | |
| VHALNNVNK | |
| 504.7828->474.2665 | |
| 504.7828->588.3094 | |
| 504.7828->701.3935 | |
| 504.7828->772.4306 | |
| VHALNNVNK[13C6] | |
| 507.7929->480.2867 | |
| 507.7929->594.3297 | |
| 507.7929->707.4137 | |
| 507.7929->778.4508 | |

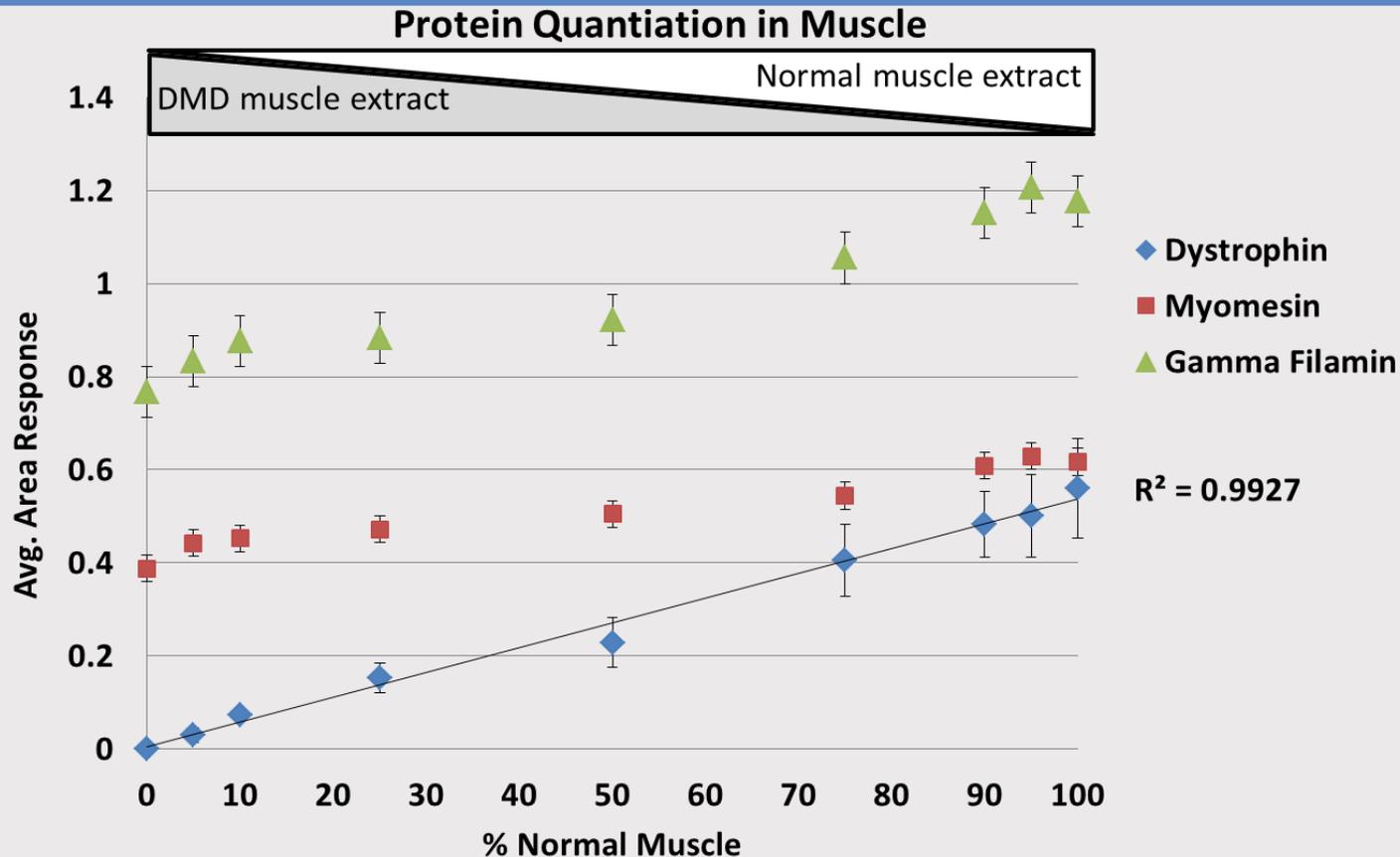


Dystrophin Peptide Quantitation w ith Increasing amount of Normal Muscle



Area response calculated by summing area across all gel slices.

Quantification of Dystrophin Protein by Mass Spectrometry

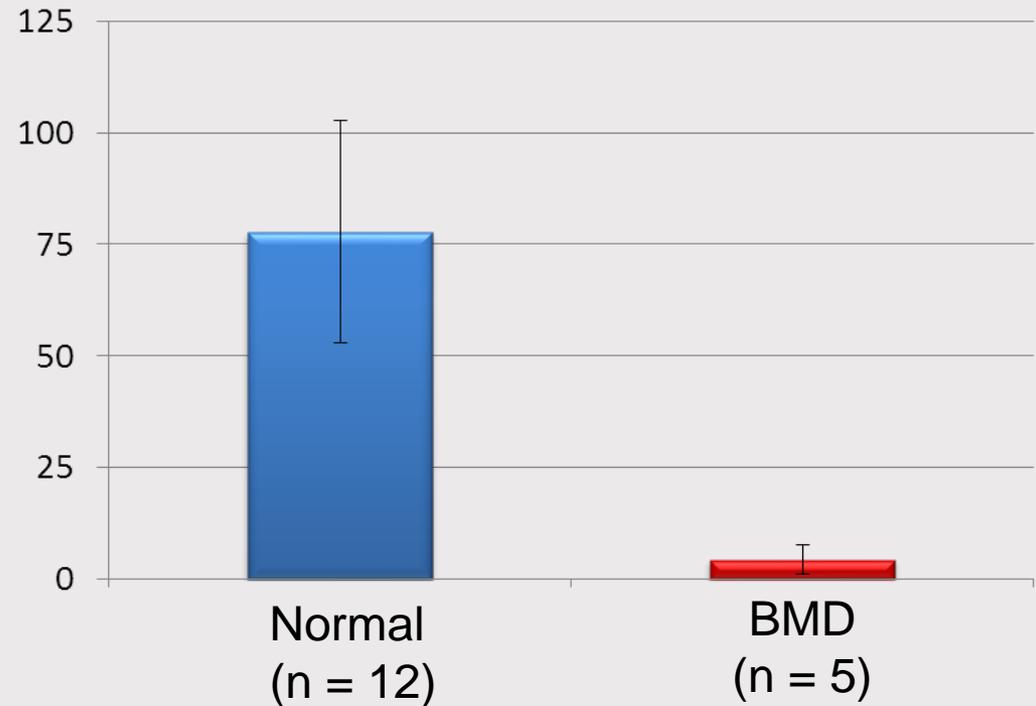


- Linear response of DMD from 0 to 100%, detecting as low as 5% of Normal
- Muscle specific proteins serve as internal standards, with level response

Absolute Quantities of Dystrophin

| Patient | Dystrophin (ng/50 ug extract) | Dystrophin (%) |
|-------------|----------------------------------|-------------------|
| normal 02 | 7.9 | 0.002 |
| normal 06 | 3.8 | 0.001 |
| normal 07 | 16.8 | 0.003 |
| normal 08 | 18.4 | 0.004 |
| normal 11 | 11.77 | 0.002 |
| normal 14 | 5.12 | 0.001 |
| normal 18 | 13.8 | 0.003 |
| normal 25 | nd | nd |
| normla 3071 | 5.3 | 0.001 |
| normal 3154 | nd | nd |
| normal 3233 | 3.07 | 0.0006 |
| normal 3377 | 9.2 | 0.0018 |
| Becker 3010 | 0.256 | 0.0001 |
| Becker 3212 | 1.02 | 0.0002 |
| Becker 3255 | 1.02 | 0.0002 |
| Becker 3479 | 1.28 | 0.0003 |
| Becker 7560 | 0 | 0.0000 |

Dystrophin (ng/mg muscle)



* Values are normalized to filamin C

FDA: Efficacy Surrogate Endpoint

Biomarker: “a characteristic that is objectively measured and evaluated as an indicator of normal biological processes, pathogenic processes, or pharmacologic responses to an intervention.” Example: cholesterol level.

Surrogate endpoint Biomarker: Biomarkers used for phase III clinical testing and biomarkers used to substantiate claims for product

“idea of a surrogate endpoint is to enable faster, smaller, more efficient clinical trials that can address urgent needs and facilitate the advancement of medicine”

Dr. Robert Temple of the Center for Drug Evaluation and Research (CDER)

http://www.nap.edu/catalog.php?record_id=12869

CDER Biomarker Qualification Program

- Pre-Submission {
 1. Conference call (**our current stage**)
 2. Cover Letter
- Consultation & Advice {
 3. Letter of Intent
 4. Qualification Review Team (QRT) Formed
 5. Initial Briefing Package Submitted
 6. Meeting with QRT
 7. Briefing Documents submitted with revisions
- Full Review {
 8. Qualification Package Submitted
 9. Formal review by QRT and CDER
 10. Final Decision

<http://www.fda.gov/Drugs/DevelopmentApprovalProcess/DrugDevelopmentToolsQualificationProgram/ucm284076.htm>



Summary/Questions

- Dystrophin is a relevant surrogate biomarker for phase 2 trials aimed at restoring dystrophin expression.
- All dystrophin restoration therapies to date aim to produce abnormal dystrophin in patient muscle, not normal dystrophin
 - Stop codon read through: amino acid replacements
 - Exon skipping: internally deleted “Becker-like” dystrophin
 - Gene therapy: dystrophin mini-gene constructs

Summary/Questions

- Abnormal dystrophin clearly therapeutic in animal models.
- Abnormal dystrophin clearly mitigates phenotype in Becker patients.
- Extensive genotype/phenotype studies suggest ~5%-10% needed:
 - Female skewed X inactivation patterns
 - Becker muscular dystrophy patients
 - Mouse models
- Is dystrophin testing of muscle a good surrogate biomarker to take through qualification process?